Supplementary Methods

Preparation of phosphorothioate-containing *HIS4* **DNA templates** Phosphorothioatecontaining templates in which phosphorothioate is incorporated into the non-template strand were prepared by ligating the products of two PCR reactions. The first PCR reaction using pSH515¹ as the template and upstream primer p965 (5'-biotin-TAATGCAGCTGGCACGACAGG-3') and downstream primer p515NTBio (5'-TCCG<u>CAGCGACTG</u>AGCATACTACTGTTCTCGAGGT-3') resulted in the production of a 356-bp fragment. The second PCR reaction with pSH515 as the template and upstream (phosphorothioate-containing) primer p515NT1 (5'-

AGGCCAGTCGCTGGTGTATATaAtAgCtAtGGAACGTTCGATTCACCTCCGATGT GTGTTGTACATACATAAAAATATCATAGCACAAC-3') or various primers with the 5 phosphorothioate bases in different positions (see Fig. 1a) and downstream primer pNOT (5'-GGCCGCTCTAGCTGCATTAATG-3') resulted in the production of a 234bp fragment. The AlwNI restriction site is underlined and lower case letters highlight the positions of the phosphorothioate bases. Each PCR product was purified using a Purelink PCR purification kit (Invitrogen) and digested with AlwNI (New England Biolabs) overnight at 37°C. The digested PCR product was then purified again using a Purelink PCR purification kit (Invitrogen). AlwNI digested products were then ligated in a reaction containing 10 µg of 356-bp product and 7 µg of 234-bp product, 66 mM Tris HCl, pH 7.5, 10 mM MgCl₂, 1 mM DTT, 1 mM ATP, 7.5% (w/v) PEG₈₀₀₀ and 2000 units of T4 DNA Ligase (NEB), overnight at room temperature. The ligation was then ethanol precipitated, washed in 80% (v/v) ethanol and the DNA pellet was dried and resuspended in 20 μ l of H₂0. Ligated DNA was size-separated on a 1% (w/v) agarose gel for 80 minutes and the 590-bp product was excised and gel purified using a QIAquick gel extraction kit (Qiagen). The biotinylated phosphorothioate-containing HIS4 promoter DNA was then used in FeBABE conjugation reactions. Phosphorothioate-containing templates with phosphorothioate incorporated into the template strand were prepared using a similar strategy.

1

Rad25 Structure Modeling and Model Evaluation For structure modeling, the helicase domains of S. cerevisiae Rad25 were aligned with Archaeoglobus fulgidus XPB. For this alignment, ClustalX was first used to align Af XPB and the two helicase subdomains of human XPB, incorporating secondary structure constraints from Af XPB (2FWR)². Separately, ClustalX was used to align sequences of eukaryotic XPBs including yeast Rad25. These two sequence alignments were then aligned using ClustalX. This initial alignment was used in conjunction with the program Modeller $8v2^3$ to generate 30 initial models. These initial models were evaluated using statistically based structure validation methods⁴⁻⁶. This analysis revealed several localized problem areas that primarily corresponded to short segments of Rad25 that weakly aligned with the Af XPB sequence. The DEVH domain of the Rad25/Af XPB alignment was next adjusted based on the NCBI alignment of the SSL2 helicase family (COG1061). This adjustment of the alignment, in conjunction with two additional rounds of modeling, resulted in several structure models which were all very similar and scored reasonably well in statistical validation tests. The final sequence alignment used for modeling is listed in Supplementary Figure 6. The two helicase domains shown in Figure 5b are the DNA damage recognition domain + N-terminal helicase subdomain (including the DEVH motif) and the C-terminal helicase subdomain. The Rad25 N-terminal domain model was based on modeling of Rad25 residues 294-714 with the full length Af XPB structure (2FWR) and truncating the model at the site of the flexible hinge region (Rad25 residue 538). The Rad25 C-terminal domain model was based on modeling Rad25 residues 548-714 with the C-terminal Af XPB structure (2FZL). Next, these two structure models were positioned in the closed orientation as described^{2,7} by superimposing the models with the HCV RNA helicase structure containing single stranded RNA (1HEI).

Yeast Strains, Nuclear Extracts and Antisera The following yeast strains were used for this study: BY4705⁸, wildtype (all strains are derivatives of this); SHY551 and 552 containing a triple Flag epitope at the C terminus of Rpb1 and Rpb2, respectively; SHY485 containing a triple Flag epitope at the C terminus of Rad25; SHY384 and 564 containing a triple Flag epitope at the C terminus of Tfg1 and Tfg2, respectively;

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SHY772 containing a 12 Flag epitope at the C terminus of Tfg1; SHY361 and 560 containing a triple Flag epitope at the C terminus of Tfa1 and Tfa2, respectively; SHY386 containing a triple Flag epitope at the C terminus of Sua7. Yeast nuclear extracts were made as described (<u>www.fhcrc.org/labs/hahn</u>). Anti-Flag M2 monoclonal antibody was from Sigma, and rabbit polyclonal antisera was generated against the N-terminal 200 amino acids of *S. cerevisiae* Rpb1⁹.

Supplementary References

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PROMOTER	Rpb1 Int residue	Rpb2 Int	Rad25 In residue	t Tfg1 residue	Int	Tfg2 residue	Int	Tfa1 residue	Int	Tfa2 residue	Int	TFIIB residue	Int
-5													
-4				331 421	M W								
-3		180 M/S protrusion											
-2		180 M/S protrusion				292	W						
-1						292	W					core domain	W
ΤΑΤΑ				316	М								
1	262 M/S clamp core			316	M/S	292	W			224	W		
2	262 M/S clamp core			316	M/S	292	W	20	W	224	W	112 ^{linker} 156 core domair	W W
3	262 M/S clamp core	180M/Sprotrusion227IobeM/S		331	M/S	184	Μ	25	W	224	w	105 linker	W
4	262 M/S clamp core	364 M/S lobe 523 W fork		331	M/S	184	Μ	25	W			99 B-finger	W
5	214 M/S clamp head	195 P M/S 239 L M/S 364 L W 503 F W		421 528	W W	196	M/S						
6	214 M/S clamp head 1187 M/S Jaw	169 P M/S 217 L M/S 532 F W	438 M/3 DEAD box domain 484 M/3	5 528	W	199	M/S						
7	214 M/S clamp head 1187 M/S Jaw		domain 627 M/3 Ct helicase domain	5									
8	1173 M/S _{Jaw}		484WDEAD box domain511511W										
9			DEAD box domain 516 W DEAD box domain								_		_
10													
11													

Supplementary Table 1 Miller and Hahn

Supplementary Table 1. Summary of RNA Polymerase II and General Transcription Factor cleavage sites generated by FeBABE attached to *HIS4* promoter DNA The measured residue where hydroxyl radical cleavage is observed is listed as well as the structural element, if known, where this residue is situated. Int., cleavage intensity. s, strong; m, medium; w, weak. P, protrusion; L, lobe; F, fork.

Supplementary Figure 1 Miller and Hahn



Supplementary Figure 1. Directed hydroxyl radical probing of RNA Polymerase II subunits Rpb1 and Rpb2 with FeBABE attached to the template strand (a) 11 FeBABE-tethered probes derived from the *HIS4* promoter where FeBABE is attached to the template strand. The color scheme is the same as in Figure 1. (b) and (c) Cleavage fragments of Rpb1-Flag and of Rpb2-Flag were visualized by Western Blotting with anti-Flag antibody as described in **Figure 2**.









Supplementary Figure 2. In vitro translated peptide fragments of Rpb1 and Rpb2 used as molecular standards for calculating FeBABE cleavage sites (a, c and e) In vitro translated peptide fragments of the N-terminal region of Rpb1-Flag, the C-terminal region of Rpb1-Flag and the N-terminal region of Rpb2-Flag, respectively, analyzed by SDS-PAGE. The residue number indicates either the last (a) or first (c and e) residue for each individual peptide fragment. (b, d and f) Calibration curves for determining the amino acid numbers of FeBABE cleavage fragments. Each calibration curve is derived from a 4th order polynomial function using the sizes (log Da (mw)) of in vitro translated peptides and their corresponding residue numbers in (a), (c) and (e). Locations of the FeBABE cleavage fragments on the calibration curves are shown with red triangles.

Supplementary Figure 3 Miller and Hahn



Supplementary Figure 3 contd. Miller and Hahn



Supplementary Figure 3 contd. Miller and Hahn



Supplementary Figure 3. In vitro translated peptide fragments of Sua7, Rad25, Tfg1, Tfg2, Tfa1 and Tfa2 used as molecular standards for calculating FeBABE cleavage sites (a, c, e, g, i and k) In vitro translated peptide fragments of the N-terminal regions of Sua7-Flag (a), Tfg2-Flag (g) and Tfa1-Flag (i) and the C-terminal regions of Rad25-Flag (c), Tfg1-Flag (e) and Tfa2-Flag (k) analyzed by SDS-PAGE. The residue number indicates the last residue for each individual peptide fragment. (b, d, f, h, j, and I) Calibration curves for determining the amino acid numbers of FeBABE cleavage fragments, similar to those shown in Supplementary Figure2.

Supplementary Figure 4 Miller and Hahn



Supplementary Figure 4. Directed hydroxyl radical probing of TFIIB in the PIC (a) Cleavage fragments of TFIIB were visualized by Western Blotting with anti-Flag antibody where FeBABE is either attached to the non-template strand or (b) template strand. Arrow points to full length TFIIB and specific cleavage fragments are indicated by the brackets and labeled as to which region in the protein is cleaved. * Indicates non-specific bands detected by antibodies. (c) Schematic indicating TFIIB domains. Arrows indicate cleavage sites.

Supplementary Figure 5 Miller and Hahn



Supplementary Figure 5. Directed hydroxyl radical probing of TFIIH subunit Rad25 and TFIIF subunits Tfg1 and Tfg2 with FeBABE attached to the template strand.

(a-c), Cleavage fragments of Rad25-Flag, Tfg1-Flag and Tfg2-Flag were visualized by Western blotting with anti-Flag antibody, respectively. Same assay as described in Figure 2. * Indicates non-specific bands detected by antibodies.

Supplementary Figure 6 Miller and Hahn ISS AF XPB bBBBb....bBBBb....aaaAAAaaa AF XPB MOMIAEIYYERGTIVVKGDAHVP--HAKFDSRSGTYRALAFRYRDIIEYFE IPKDLFDFYEQMDKDEEEEEET--QTVSFEVKQEMIEELQKRCIC-LEYPL mouseXPB IPMDLFDFYEQMDKDEEEEEET--QTVSFEVKQEMIEELQKRCIH-LEYPL humanXPB VPEDITDFYEKIDKEEEDEDEANLKTVSFEVAQEKIEVIQKRCIE-IEHPL drosXPB VFSAVIGGDNEREEEDDDIDAVH----SFEIANESVEVVKKRCQE-IDYPV Scerev_rad25 SpombeXPB LFSAVVG----LQEEEDDEDAVH----LFEIKHSSVETIKKRCAE-IDYPL *: : . . . : : ::: ISS AF XPBbBBBb......bBBb.aaaAAAAAaaa...--bBBBb.....aaaAA AF XPB SNGIEFVDNAADPIPTPYFDAEISLRDYQEKALERWLVDKR---GCIVLPTGSGKTHVAM mouseXPB LAEYDFRNDTLNPDINIDLKPTAVLRPYQEKSLRKMFGNGRARSGVIVLPCGAGKSLVGV humanXPB LAEYDFRNDSVNPDINIDLKPTAVLRPYQEKSLRKMFGNGRARSGVIVLPCGAGKSLVGV drosXPB LAEYDFRNDTNNPDINIDLKPAAVLRPYQEKSLRKMFGNGRARSGVIVLPCGAGKSLVGV Scerev_rad25 LEEYDFRNDHRNPDLDIDLKPSTQIRPYQEKSLSKMFGNGRARSGIIVLPCGAGKTLVGI SpombeXPB LEEYDFRNDNINPDLPIDLKPSTQIRPYQEKSLSKMFGNGRARSGIIVLPCGAGKTLVGI :* ****:* : : : : * **** *:**: *.: :* :: :* :.. ISS AF XPB AAAaaa...bBBBb..aaaAAAAaaa.....bBBb..bBBb.-..-bBBBB AAINELSTPTLIVVPTLALAEOWKERLGIFG---EEYVGEFSGRIKELKP----LTVSTY AF XPB mouseXPB TAACTVRKRCLVLGNSAVSVEQWKAQFKMWSTIDDSQICRFTSDAKD-KPIGCSVAISTY TAACTVRKRCLVLGNSAVSVEQWKAQFKMWSTIDDSQICRFTSDAKD-KPIGCSVAISTY humanXPB drosXPB TACCTVRKRALVLCNSGVSVEQWKQQFKMWSTADDSMICRFTSEAKD-KPMGCGILVTTY Scerev_rad25 TAACTIKKSVIVLCTSSVSVMQWRQQFLQWCTLQPENCAVFTSDNKEMFQTESGLVVSTY SpombeXPB TAACTIKKSVIVLCTSSVSVMQWRQQFLQWSNIKPDHIAVFTADHKERFHSEAGVVVSTY :* : . ::: : . . **: :: : * . * . . ISS AF XPB b-----aaaAAAaaa----...bBBBb.....aaaaaa....bBBBBb... D-----SAYVNAEKL----GNRFMLLIFDEVHHLPAESYVQIAQMSIAPFRLGLTATF AF_XPB mouseXPB SMLGHTTKRSWEAERVMEWLKTQEWGLMILDEVHTIPARMFRRVLTIVQAHCKLGLTATL humanXPB SMLGHTTKRSWEAERVMEWLKTOEWGLMILDEVHTIPAKMFRRVLTIVOAHCKLGLTATL SMITHTQKRSWEAEQTMRWLQEQEWGIMVLDEVHTIPAKMFRRVLTIVQSHCKLGLTATL drosXPB Scerev_rad25 SMVANTRNRSHDSQKVMDFLTGREWGFIILDEVHVVPAAMFRRVVSTIAAHAKLGLTATL SMVANTRNRSYDSQKMMDFLTGREWGFILLDEVHVVPAAMFRRVVTTIAAHTKLGLTATL SpombeXPB ••• ••• : :: : :: ****** !SS_AF_XPBaaaAaaa...bBBBb.....bBBBBBb..... AF XPB EREDGRHEILKEVVGGKVFELFPDSLAG-KHLAKYTIKRIFVPLAEDERVEYEKREKVYK mouseXPB VREDDKIVDLNFLIGPKLYEANWMELQNNGYIAKVQCAEVWCPMSPEFYREYVAIKTKKR humanXPB VREDDKIVDLNFLIGPKLYEANWMELONNGYIAKVOCAEVWCPMSPEFYREYVAIKTKKR drosXPB LREDDKIADLNFLIGPKLYEANWLELQKKGYIARVQCAEVWCPMSPEFYREYLTTKTSKK VREDDKIGDLNFLIGPKLYEANWMELSQKGHIANVQCAEVWCPMTAEFYQEYLRETARKR Scerev rad25 SpombeXPB VREDDKIDDLNFLIGPKMYEANWMDLAQKGHIAKVQCAEVWCAMTTEFYNEYLRENSRKR ***.: *: ::* *::* : ::*. •* : !SS_AF_XPB AF XPB QFLRARGITLRRAEDFNKIVMASGYDERAYEALRAWEEARRIAFNSKNKIRKLREILERH mouseXPB ILLYTMN-----PNKFRACQFLIKFHER ILLYTMN-----PNKFRACQFLIKFHER humanXPB MLLYVMN-----PSKFRSCQFLIKYHEQ drosXPB MLLYIMN-----PTKFOACOFLIQYHER Scerev rad25 MLLYIMN-----PKKFQACQFLIDYHEK SpombeXPB :* .* : :: . . : ...bBBBb.....aaaAAaaa...bBBBb.....aaaAaaa...-.bBBBBBBb.... !SS_AF_XPB AF XPB RKDKIIIFTRHNELVYRISKVFLIPAITHRTSREEREEILEGFRTG-RFRAIVSSQVLDE mouseXPB RNDKIIVFADNVFALKEYAIRLNKPYIYGPTSQGERMQILQNFKHNPKINTIFISKVGDT RNDKIIVFADNVFALKEYAIRLNKPYIYGPTSQGERMQILQNFKHNPKINTIFISKVGDT humanXPB drosXPB RGDKTIVFSDNVFALKHYAIKMNKPFIYGPTSQNERIQILQNFKFNSKVNTIFVSKVADT Scerev rad25 RGDKIIVFSDNVYALQEYALKMGKPFIYGSTPOQERMNILQNFQYNDQINTIFLSKVGDT RGDKIIVFSDNVYALRAYAIKLGKYFIYGGTPQQERMRILENFQYNELVNTIFLSKVGDT SpombeXPB *.: ** .**:.*: . * ** *:*: : : : : * ...*. *:* *bBBBb...-..bBBBbb..... !SS_AF_XPB AF_XPB GIDVPDANVGVIMSGSG-SAREYIQRLGRILRPSKG----KKEAVLYELISRGTG mouseXPB SFDLPEANVLIOISSHGGSRROEAORLGRVLRAKKGMVAEEYNAFFYSLVSODTO SFDLPEANVLIQISSHGGSRRQEAQRLGRVLRAKKGMVAEEYNAFFYSLVSQDTQ humanXPB drosXPB SFDLPEANVLIQISSHGGSRRQEAQRLGRILRAKKGAIAEEYNAFFYTLVSQDTM Scerev rad25 SIDLPEATCLIQISSHYGSRRQEAQRLGRILRAKR-RNDEGFNAFFYSLVSKDTQ SIDLPEATCLIQISSHYGSRRQEAQRLGRILRAKR-RNDEGFNAFFYSLVSKDTQ SpombeXPB .:*:*:*. : :*. * *: ****:**..: :*.:* *:*:.*

Supplementary Figure 6. Rad25/XPB alignment used in structure modeling of the Rad25 helicase domains

Rad25/XPB sequence alignment used in structure modeling of the Rad25 helicase domains. Shown is the alignment of the Rad25/XPB helicase subdomains with Archaeoglobus fulgidus XPB (Af_XPB). Shown at the top is the secondary structure of the Af XPB structure (2FWR). The alignment was derived as described in **Supplementary Methods**.

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Supplementary Figure 1 Miller and Hahn



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Supplementary Figure 3 Miller and Hahn



Supplementary Figure 3 contd. Miller and Hahn



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Supplementary Figure 3. In vitro translated peptide fragments of Sua7, Rad25, Tfg1, Tfg2, Tfa1 and Tfa2 used as molecular standards for calculating FeBABE cleavage sites (a, c, e, g, i and k) In vitro translated peptide fragments of the N-terminal regions of Sua7-Flag (a), Tfg2-Flag (g) and Tfa1-Flag (i) and the C-terminal regions of Rad25-Flag (c), Tfg1-Flag (e) and Tfa2-Flag (k) analyzed by SDS-PAGE. The residue number indicates the last residue for each individual peptide fragment. (b, d, f, h, j, and I) Calibration curves for determining the amino acid numbers of FeBABE cleavage fragments, similar to those shown in Supplementary Figure2.

Supplementary Figure 4 Miller and Hahn



Supplementary Figure 4. Directed hydroxyl radical probing of TFIIB in the PIC (a) Cleavage fragments of TFIIB were visualized by Western Blotting with anti-Flag antibody where FeBABE is either attached to the non-template strand or (b) template strand. Arrow points to full length TFIIB and specific cleavage fragments are indicated by the brackets and labeled as to which region in the protein is cleaved. * Indicates non-specific bands detected by antibodies. (c) Schematic indicating TFIIB domains. Arrows indicate cleavage sites.

Supplementary Figure 5 Miller and Hahn



Supplementary Figure 5. Directed hydroxyl radical probing of TFIIH subunit Rad25 and TFIIF subunits Tfg1 and Tfg2 with FeBABE attached to the template strand.

(a-c), Cleavage fragments of Rad25-Flag, Tfg1-Flag and Tfg2-Flag were visualized by Western blotting with anti-Flag antibody, respectively. Same assay as described in Figure 2. * Indicates non-specific bands detected by antibodies.

Supplementary Figure 6 Miller and Hahn ISS AF XPB bBBBb....bBBBb....aaaAAAaaa AF XPB MOMIAEIYYERGTIVVKGDAHVP--HAKFDSRSGTYRALAFRYRDIIEYFE IPKDLFDFYEQMDKDEEEEEET--QTVSFEVKQEMIEELQKRCIC-LEYPL mouseXPB IPMDLFDFYEQMDKDEEEEEET--QTVSFEVKQEMIEELQKRCIH-LEYPL humanXPB VPEDITDFYEKIDKEEEDEDEANLKTVSFEVAQEKIEVIQKRCIE-IEHPL drosXPB VFSAVIGGDNEREEEDDDIDAVH----SFEIANESVEVVKKRCQE-IDYPV Scerev_rad25 SpombeXPB LFSAVVG----LQEEEDDEDAVH----LFEIKHSSVETIKKRCAE-IDYPL *: : . . . : : ::: ISS AF XPBbBBBb......bBBb.aaaAAAAAaaa...--bBBBb.....aaaAA AF XPB SNGIEFVDNAADPIPTPYFDAEISLRDYQEKALERWLVDKR---GCIVLPTGSGKTHVAM mouseXPB LAEYDFRNDTLNPDINIDLKPTAVLRPYQEKSLRKMFGNGRARSGVIVLPCGAGKSLVGV humanXPB LAEYDFRNDSVNPDINIDLKPTAVLRPYQEKSLRKMFGNGRARSGVIVLPCGAGKSLVGV drosXPB LAEYDFRNDTNNPDINIDLKPAAVLRPYQEKSLRKMFGNGRARSGVIVLPCGAGKSLVGV Scerev_rad25 LEEYDFRNDHRNPDLDIDLKPSTQIRPYQEKSLSKMFGNGRARSGIIVLPCGAGKTLVGI SpombeXPB LEEYDFRNDNINPDLPIDLKPSTQIRPYQEKSLSKMFGNGRARSGIIVLPCGAGKTLVGI :* ****:* : : : : * **** *:**: *.: :* :: :* :.. ISS AF XPB AAAaaa...bBBBb..aaaAAAAaaa.....bBBb..bBBb.-..-bBBBB AAINELSTPTLIVVPTLALAEOWKERLGIFG---EEYVGEFSGRIKELKP----LTVSTY AF XPB mouseXPB TAACTVRKRCLVLGNSAVSVEQWKAQFKMWSTIDDSQICRFTSDAKD-KPIGCSVAISTY TAACTVRKRCLVLGNSAVSVEQWKAQFKMWSTIDDSQICRFTSDAKD-KPIGCSVAISTY humanXPB drosXPB TACCTVRKRALVLCNSGVSVEQWKQQFKMWSTADDSMICRFTSEAKD-KPMGCGILVTTY Scerev_rad25 TAACTIKKSVIVLCTSSVSVMQWRQQFLQWCTLQPENCAVFTSDNKEMFQTESGLVVSTY SpombeXPB TAACTIKKSVIVLCTSSVSVMQWRQQFLQWSNIKPDHIAVFTADHKERFHSEAGVVVSTY :* : . ::: : . . **: :: : * . * . . ISS AF XPB b-----aaaAAAaaa----...bBBBb.....aaaaaa....bBBBBb... D-----SAYVNAEKL----GNRFMLLIFDEVHHLPAESYVQIAQMSIAPFRLGLTATF AF_XPB mouseXPB SMLGHTTKRSWEAERVMEWLKTQEWGLMILDEVHTIPARMFRRVLTIVQAHCKLGLTATL humanXPB SMLGHTTKRSWEAERVMEWLKTOEWGLMILDEVHTIPAKMFRRVLTIVOAHCKLGLTATL SMITHTQKRSWEAEQTMRWLQEQEWGIMVLDEVHTIPAKMFRRVLTIVQSHCKLGLTATL drosXPB Scerev_rad25 SMVANTRNRSHDSQKVMDFLTGREWGFIILDEVHVVPAAMFRRVVSTIAAHAKLGLTATL SMVANTRNRSYDSQKMMDFLTGREWGFILLDEVHVVPAAMFRRVVTTIAAHTKLGLTATL SpombeXPB ••• ••• : :: : :: ****** !SS_AF_XPBaaaAaaa...bBBBb.....bBBBBBb..... AF XPB EREDGRHEILKEVVGGKVFELFPDSLAG-KHLAKYTIKRIFVPLAEDERVEYEKREKVYK mouseXPB VREDDKIVDLNFLIGPKLYEANWMELQNNGYIAKVQCAEVWCPMSPEFYREYVAIKTKKR humanXPB VREDDKIVDLNFLIGPKLYEANWMELONNGYIAKVOCAEVWCPMSPEFYREYVAIKTKKR drosXPB LREDDKIADLNFLIGPKLYEANWLELQKKGYIARVQCAEVWCPMSPEFYREYLTTKTSKK VREDDKIGDLNFLIGPKLYEANWMELSQKGHIANVQCAEVWCPMTAEFYQEYLRETARKR Scerev rad25 SpombeXPB VREDDKIDDLNFLIGPKMYEANWMDLAQKGHIAKVQCAEVWCAMTTEFYNEYLRENSRKR ***.: *: ::* *::* : ::*. •* : !SS_AF_XPB AF XPB QFLRARGITLRRAEDFNKIVMASGYDERAYEALRAWEEARRIAFNSKNKIRKLREILERH mouseXPB ILLYTMN-----PNKFRACQFLIKFHER ILLYTMN-----PNKFRACQFLIKFHER humanXPB MLLYVMN-----PSKFRSCQFLIKYHEQ drosXPB MLLYIMN-----PTKFOACOFLIQYHER Scerev rad25 MLLYIMN-----PKKFQACQFLIDYHEK SpombeXPB :* .* : :: . . : ...bBBBb.....aaaAAaaa...bBBBb.....aaaAaaa...-.bBBBBBBb.... !SS_AF_XPB AF XPB RKDKIIIFTRHNELVYRISKVFLIPAITHRTSREEREEILEGFRTG-RFRAIVSSQVLDE mouseXPB RNDKIIVFADNVFALKEYAIRLNKPYIYGPTSQGERMQILQNFKHNPKINTIFISKVGDT RNDKIIVFADNVFALKEYAIRLNKPYIYGPTSQGERMQILQNFKHNPKINTIFISKVGDT humanXPB drosXPB RGDKTIVFSDNVFALKHYAIKMNKPFIYGPTSQNERIQILQNFKFNSKVNTIFVSKVADT Scerev rad25 RGDKIIVFSDNVYALQEYALKMGKPFIYGSTPOQERMNILQNFQYNDQINTIFLSKVGDT RGDKIIVFSDNVYALRAYAIKLGKYFIYGGTPQQERMRILENFQYNELVNTIFLSKVGDT SpombeXPB *.: ** .**:.*: . * ** *:*: : : : : * ...*. *:* *bBBBb...-..bBBBbb..... !SS_AF_XPB AF_XPB GIDVPDANVGVIMSGSG-SAREYIQRLGRILRPSKG----KKEAVLYELISRGTG mouseXPB SFDLPEANVLIOISSHGGSRROEAORLGRVLRAKKGMVAEEYNAFFYSLVSODTO SFDLPEANVLIQISSHGGSRRQEAQRLGRVLRAKKGMVAEEYNAFFYSLVSQDTQ humanXPB drosXPB SFDLPEANVLIQISSHGGSRRQEAQRLGRILRAKKGAIAEEYNAFFYTLVSQDTM Scerev rad25 SIDLPEATCLIQISSHYGSRRQEAQRLGRILRAKR-RNDEGFNAFFYSLVSKDTQ SIDLPEATCLIQISSHYGSRRQEAQRLGRILRAKR-RNDEGFNAFFYSLVSKDTQ SpombeXPB .:*:*:*. : :*. * *: ****:**..: :*.:* *:*:.*

Supplementary Figure 6. Rad25/XPB alignment used in structure modeling of the Rad25 helicase domains

Rad25/XPB sequence alignment used in structure modeling of the Rad25 helicase domains. Shown is the alignment of the Rad25/XPB helicase subdomains with Archaeoglobus fulgidus XPB (Af_XPB). Shown at the top is the secondary structure of the Af XPB structure (2FWR). The alignment was derived as described in **Supplementary Methods**.